



# **Problems in Computational Biology:**

**(1) Deciphering the Protein Complex Network**

**And maybe a little bit of:**

**(2) The Shape of RNA**

**or**

**(3) Finding non-coding RNA Genes**

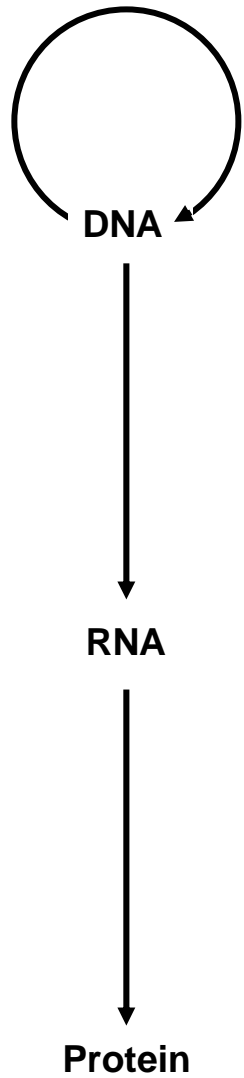
**Richard F. Meraz**

**Lawrence Berkeley National Laboratory**

**Berkeley, CA**

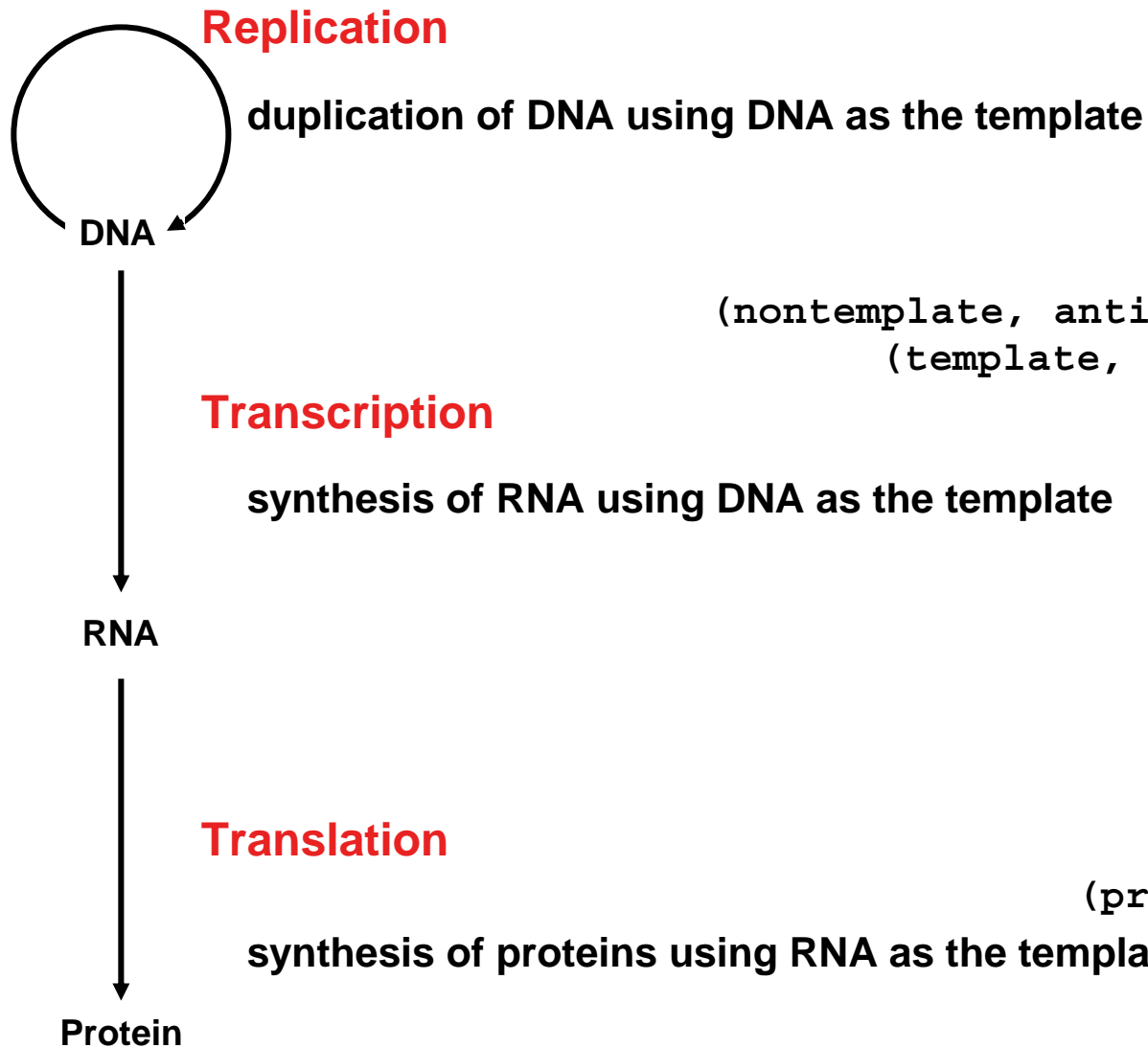


# The Central Dogma

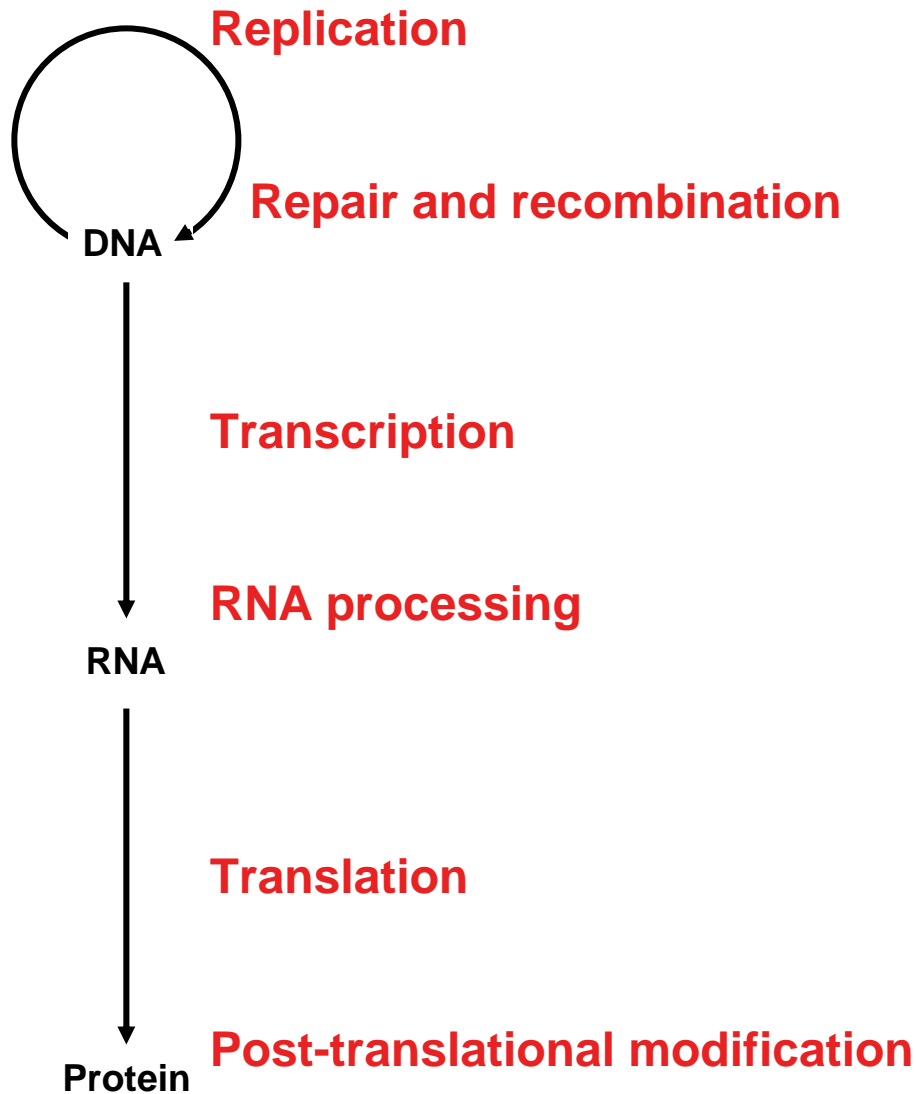




# The Central Dogma



# The Central Dogma



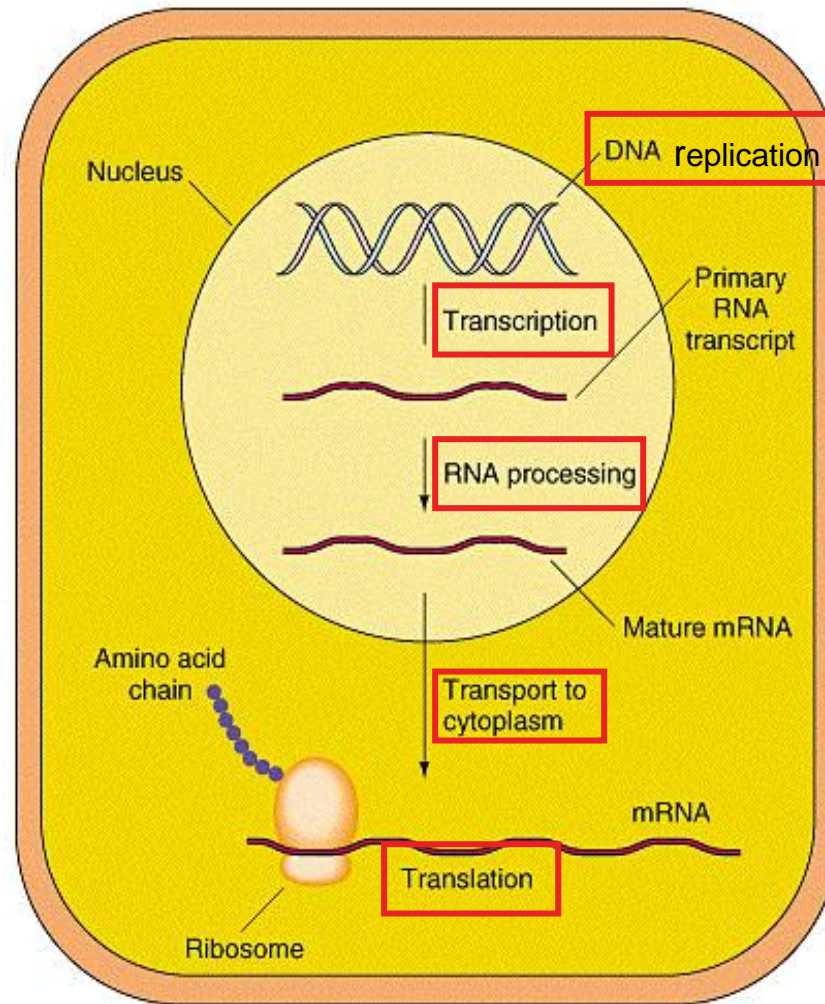
1. DNA pol  $\alpha$  and  $\delta$

2. DNA pol  $\beta$  and  $\epsilon$

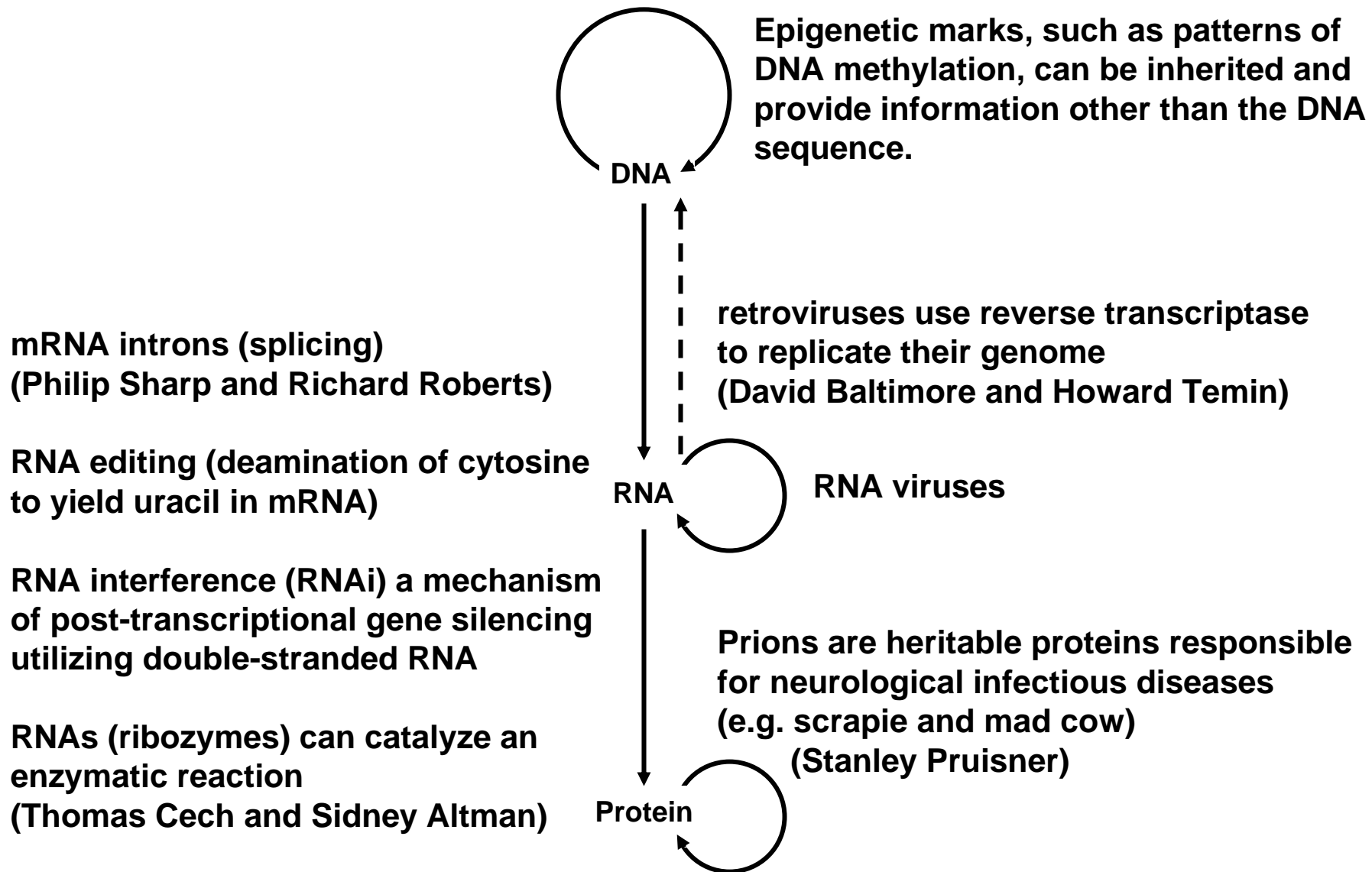
1. RNA pol I-ribosomal RNA (rRNA)
2. RNA pol II-messenger RNA (mRNA)
3. RNA pol III-5S rRNA, snRNA, tRNA

1. mRNA splicing
2. rRNA and tRNA processing
3. capping and polyadenylation

1. phosphorylation
2. methylation
3. ubiquitination

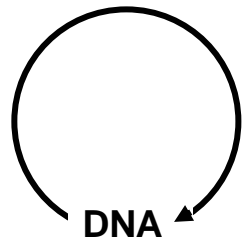


# Exceptions to the Central Dogma (get Nobel Prizes)





# The Central Dogma



Finding non-coding RNA Genes



RNA



Non-coding or functional RNA

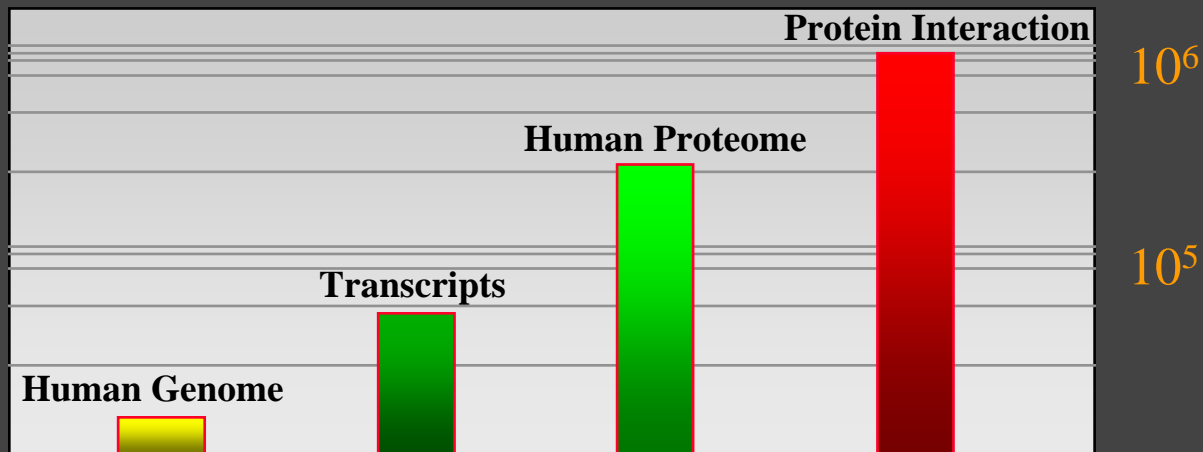


Protein

The Shape of (non-coding) RNA

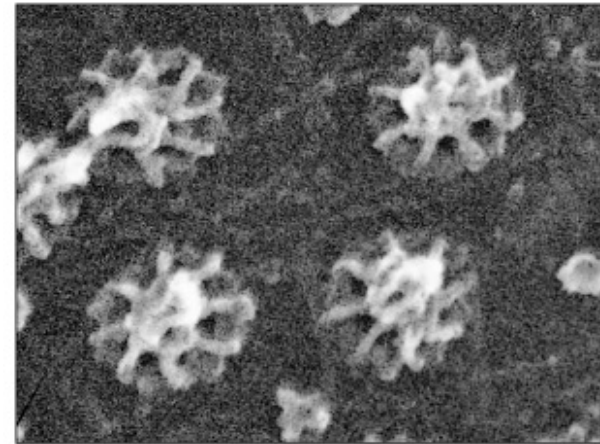
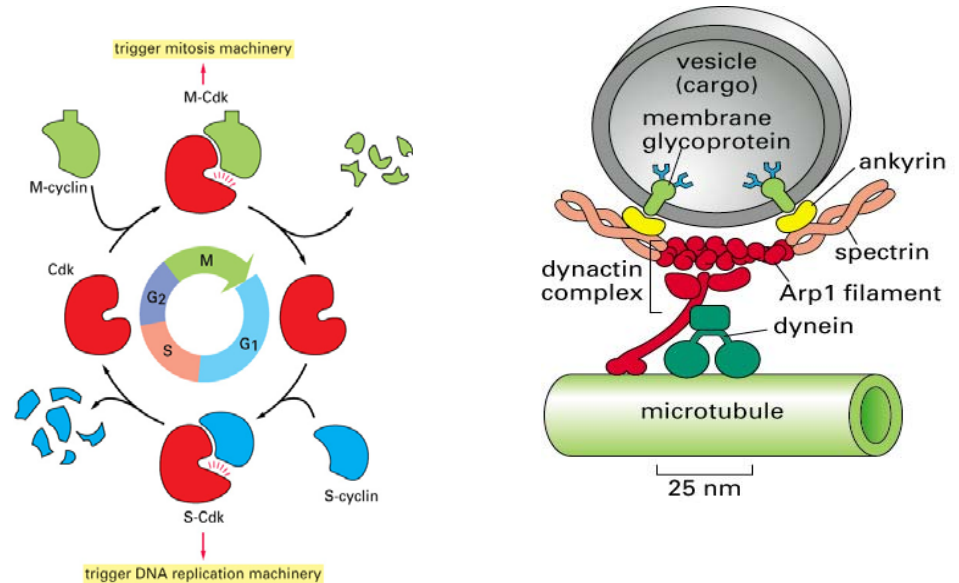
Deciphering the Protein Complex Network

Genome: 30.000 genes  
↓  
Transcriptome: 40-100.000 mRNAs  
↓  
Proteome: 100-400.000 proteins  
>1.000.000 interactions



# Protein-Complexes – Who Cares ?

- Protein complexes are important for virtually every biological process and most diseases.
- Genome sequences identify tens of thousands of genes: linking these to 200-300 core biological processes will make their study manageable.



(B)

0.1 μm



# High-throughput methods for detecting Protein Complexes

- **Two-hybrid** dataset by Uetz *et al* 2000 (the first comprehensive study in yeast)
- **Two-hybrid** dataset by Ito *et al* 2001 (broad coverage in yeast)
- **HMS-PCI** dataset by Ho *et al* 2002
- **TAP-MS** dataset by Gavin *et al* 2002

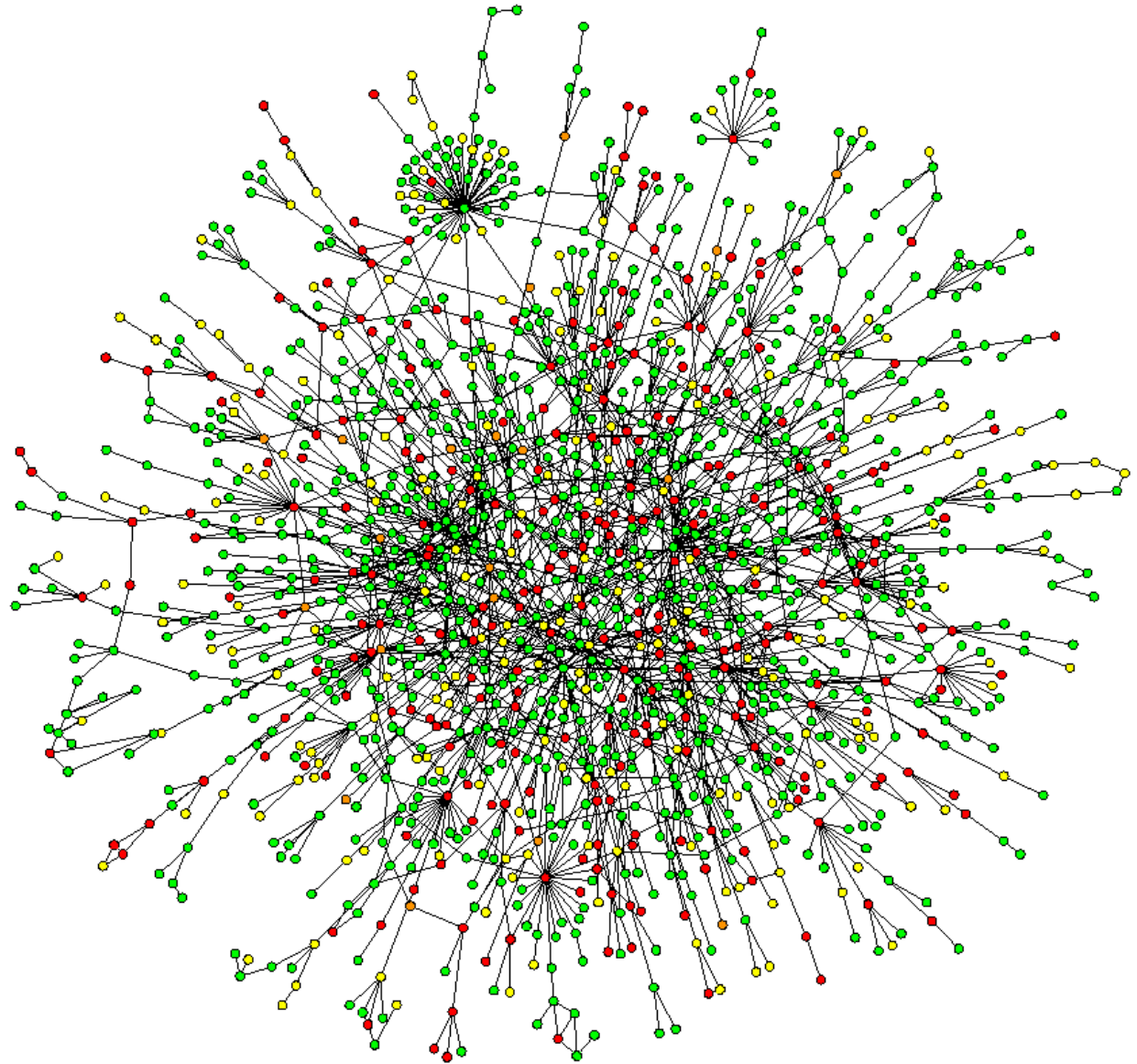


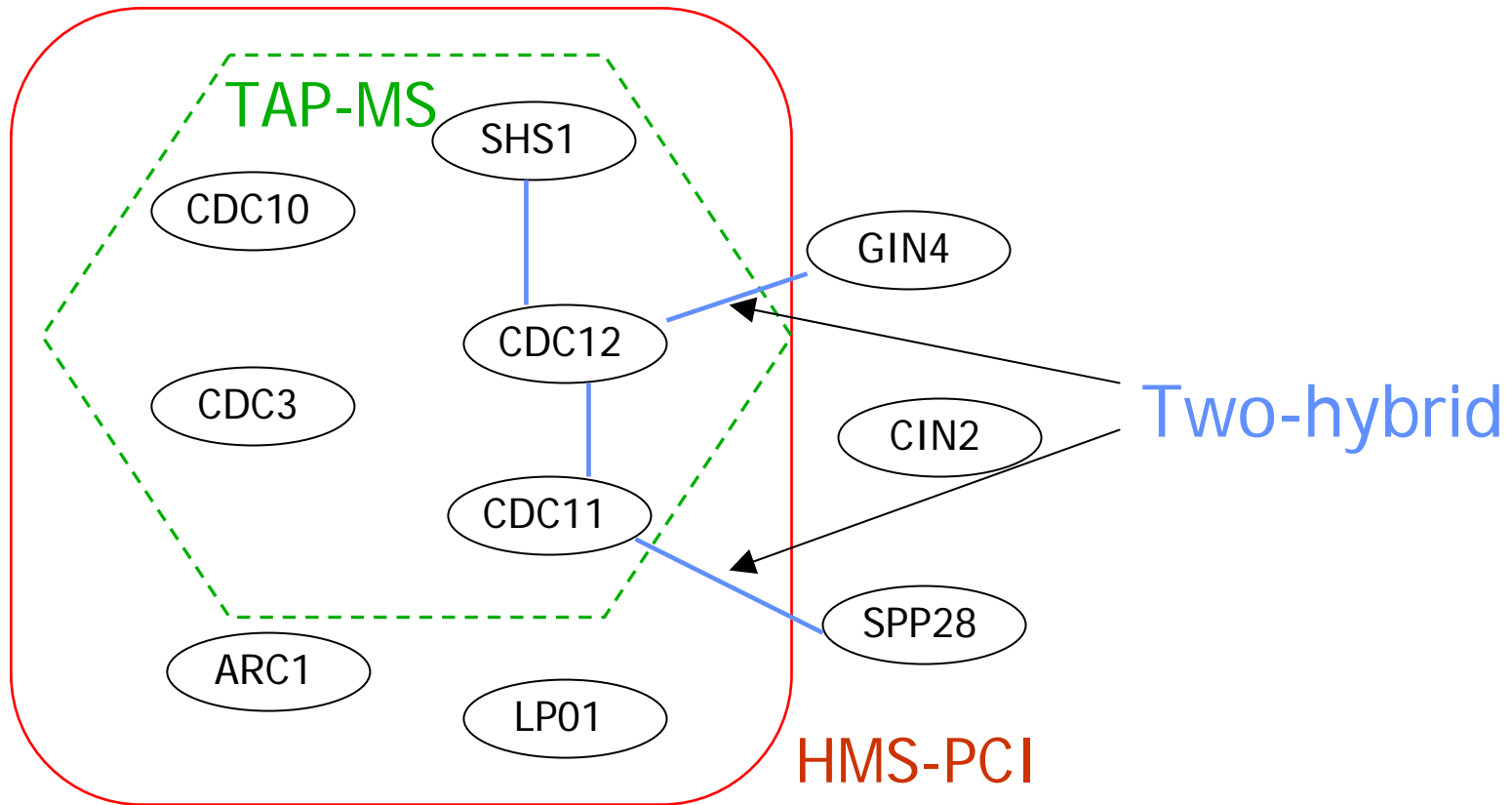
Very little overlap between interaction data from different experiments.

	<b>ITO <i>et al</i></b>	<b>Uetz <i>et al</i></b>	<b>Gavin <i>et al</i></b>	<b>Ho <i>et al</i></b>
<b>Ito <i>et al</i></b>	4363	<b>186</b>	<b>54</b>	<b>63</b>
<b>Uetz <i>et al</i></b>	<b>186</b>	1403	<b>54</b>	<b>56</b>
<b>Gavin <i>et al</i></b>	<b>54</b>	<b>54</b>	3222	<b>198</b>
<b>Ho <i>et al</i></b>	<b>63</b>	<b>56</b>	<b>198</b>	3596

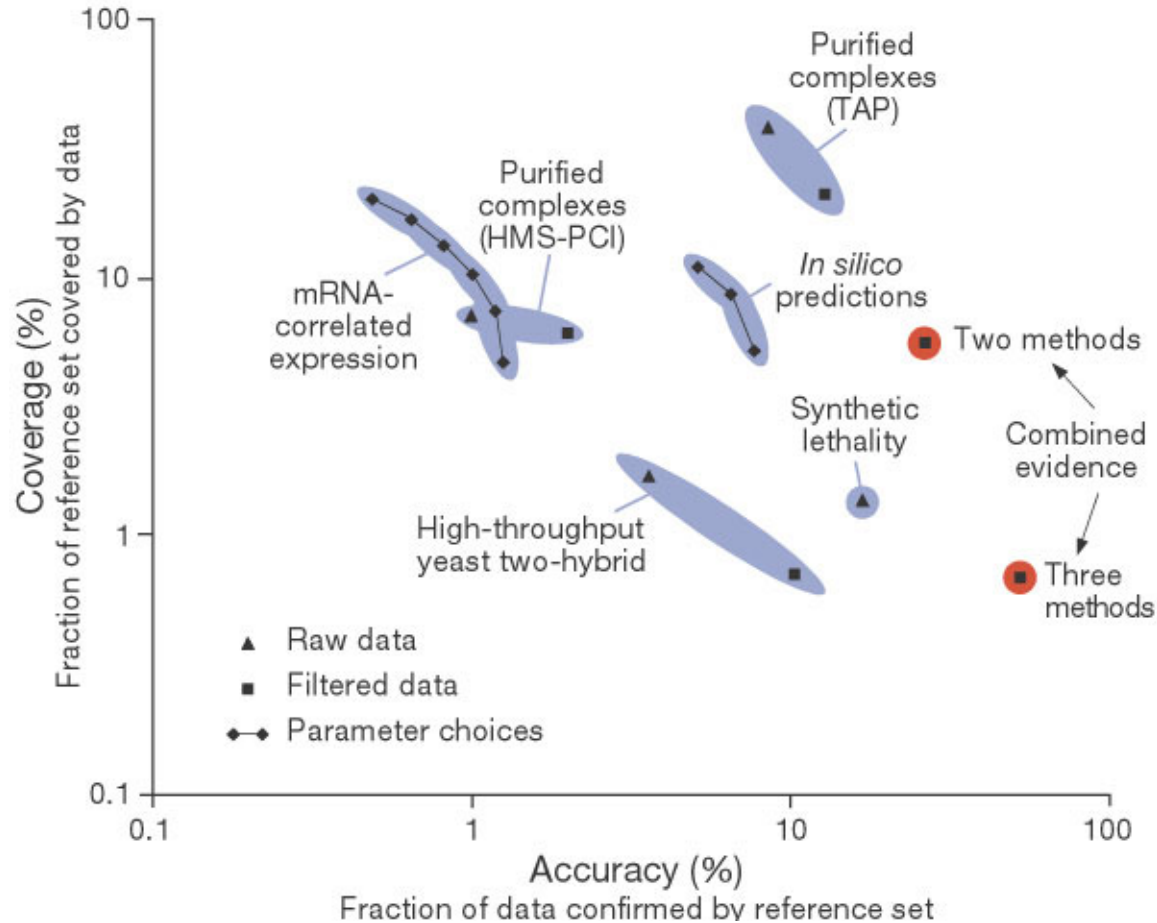
Copied from Salwinski and Eisenberg, 2003

# Interactions in the yeast proteome

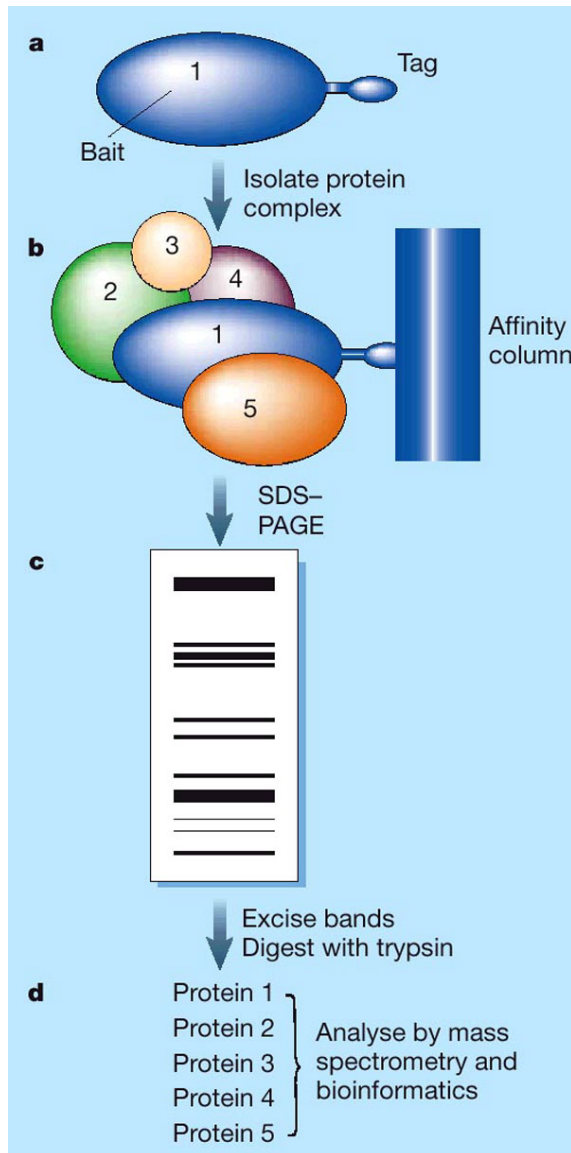




# Not all data-sets are created equal.



von Mering C, Krause R, Snel B, Cornell M, Oliver SG, Fields S, Bork P.  
Comparative assessment of large-scale data sets of protein-protein interactions.  
Nature 2002;417(6887):399-403.

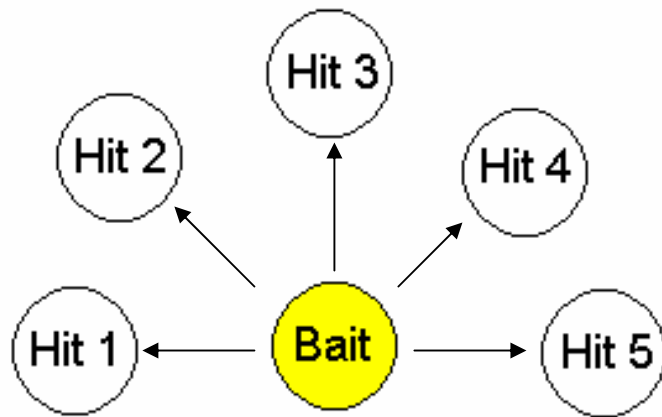


- Tandem-Affinity Purification coupled with Mass-Spectrometry (TAP-MS) determines the constituents of multi-protein complexes.

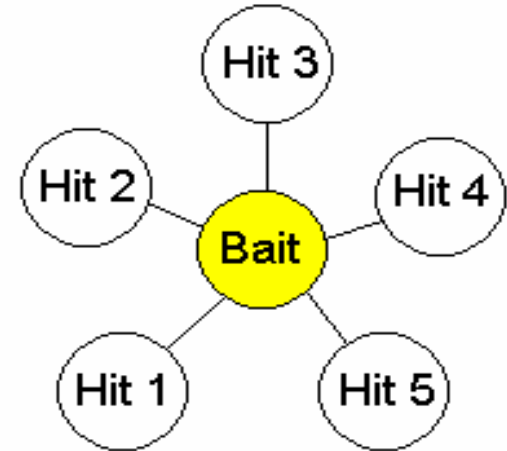
Gavin AC, *et al.* Functional organization of the yeast proteome by systematic analysis of protein complexes. *Nature* 2002;415(6868):141-147.

# Computer Scientists who don't understand biology make bad assumptions

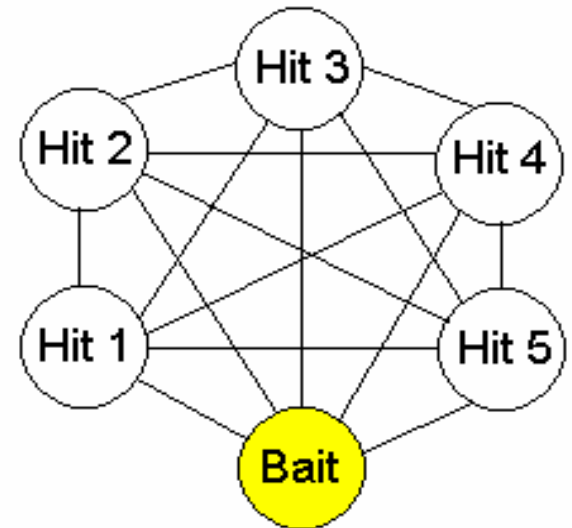
Data



Spoke  
Model



Matrix  
Model



*K*-cores (Bader and Hogue, 2002)  
Cliques (Spirin and Mirny, 2003)  
Hypergraph – *k*-core (Pothén, 2003 )



# Looking at the Network

- **Reduce noise by eliminating unnecessary assumptions about which proteins interact**
- **Do not ignore that fact that proteins that coincide in more than one complex are likely to somehow integrate their functions.**
- **Don't ignore the notion of 'communication' and coupling that occurs when protein complexes share components. This is the higher-level organization of the network via linking of biological processes.**



# Goals + Hypothesis

- **Unify two notions of network – proteins interacting, but also complexes interacting via the notion of ‘shared components’.**
- **Partition these networks into ‘modules’ or functional units separable from the rest of the network. This is the goal of systems-level or network biology.**
- **The framework should aid in reasoning about uncharacterized protein components and be biologically consistent.**



# A Unified Representation of Multi-Protein Complex Networks

**Dual** relationship between protein and protein-complex is specified by adjacency matrix **B**.

## Protein-Protein (p-p) interaction network:

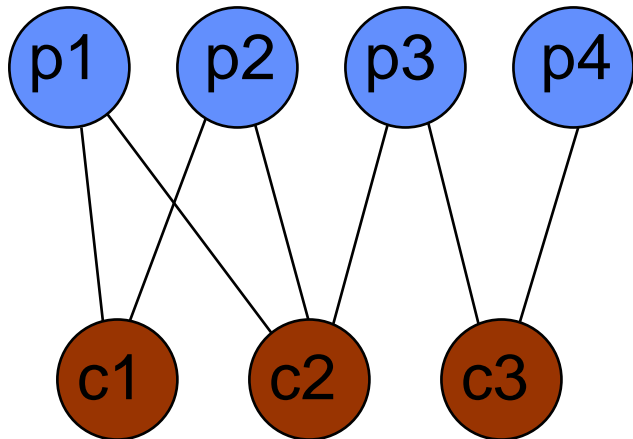
$$(\mathbf{B}\mathbf{B}^T)_{ij} = \left( \begin{array}{c} \# \text{ of protein complexes} \\ \text{containing both proteins } p_i, p_j \end{array} \right)$$

## Complex-Complex (c-c) interaction network

$$(\mathbf{B}^T\mathbf{B})_{ij} = \left( \begin{array}{c} \# \text{ of proteins shared by} \\ \text{protein complexes } c_i, c_j \end{array} \right)$$

# Toy Protein Complex Dataset

Bipartite Graph



Adjacency Matrix

$\mathbf{B}$

	c1	c2	C3
P1	1	1	0
P2	1	1	0
P3	0	1	1
P4	0	0	1

$\mathbf{B}^T$

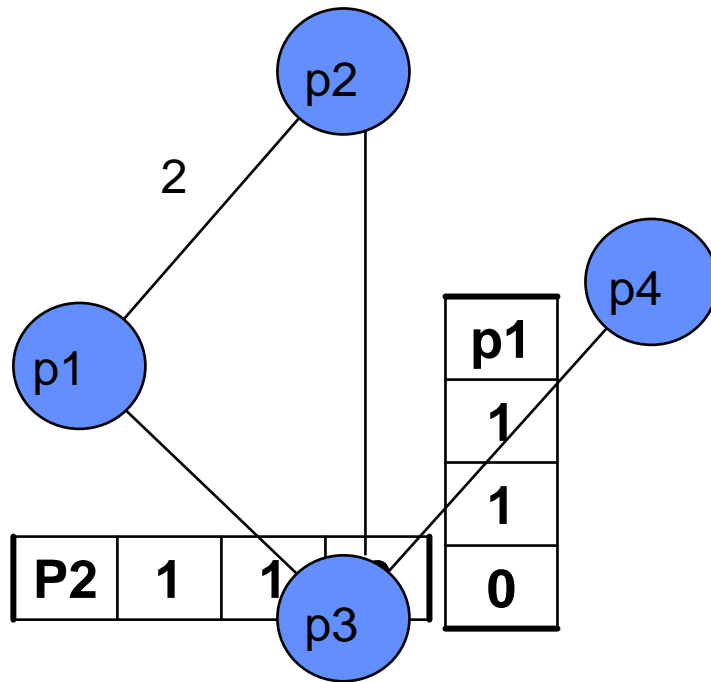
	p1	p2	p3	P4
C1	1	1	0	0
C2	1	1	1	0
C3	0	0	1	1

**B**

	c1	c2	C3
P1	1	1	0
P2	1	1	0
P3	0	1	1
P4	0	0	1

**B<sup>T</sup>**

	p1	p2	p3	P4
C1	1	1	0	0
C2	1	1	1	0
C3	0	0	1	1



	p1	p2	p3	P4
P1	2	2	1	0
P2	2	2	1	0
P3	1	1	2	1
P4	0	0	1	1



# Clustering P-P and C-C Network

$$s(G_1, G_2) = \sum_{i \in G_1} \sum_{j \in G_2} w_{ij}$$

Connectivity

$$J(G_1, G_2) = \frac{s(G_1, G_2)}{s(G_1, G_1)} + \frac{s(G_1, G_2)}{s(G_2, G_2)}$$

Cohesion between two graphs

$$q(i) = \begin{cases} a & \text{if } i \in G_1 \\ -b & \text{if } i \in G_2 \end{cases}$$

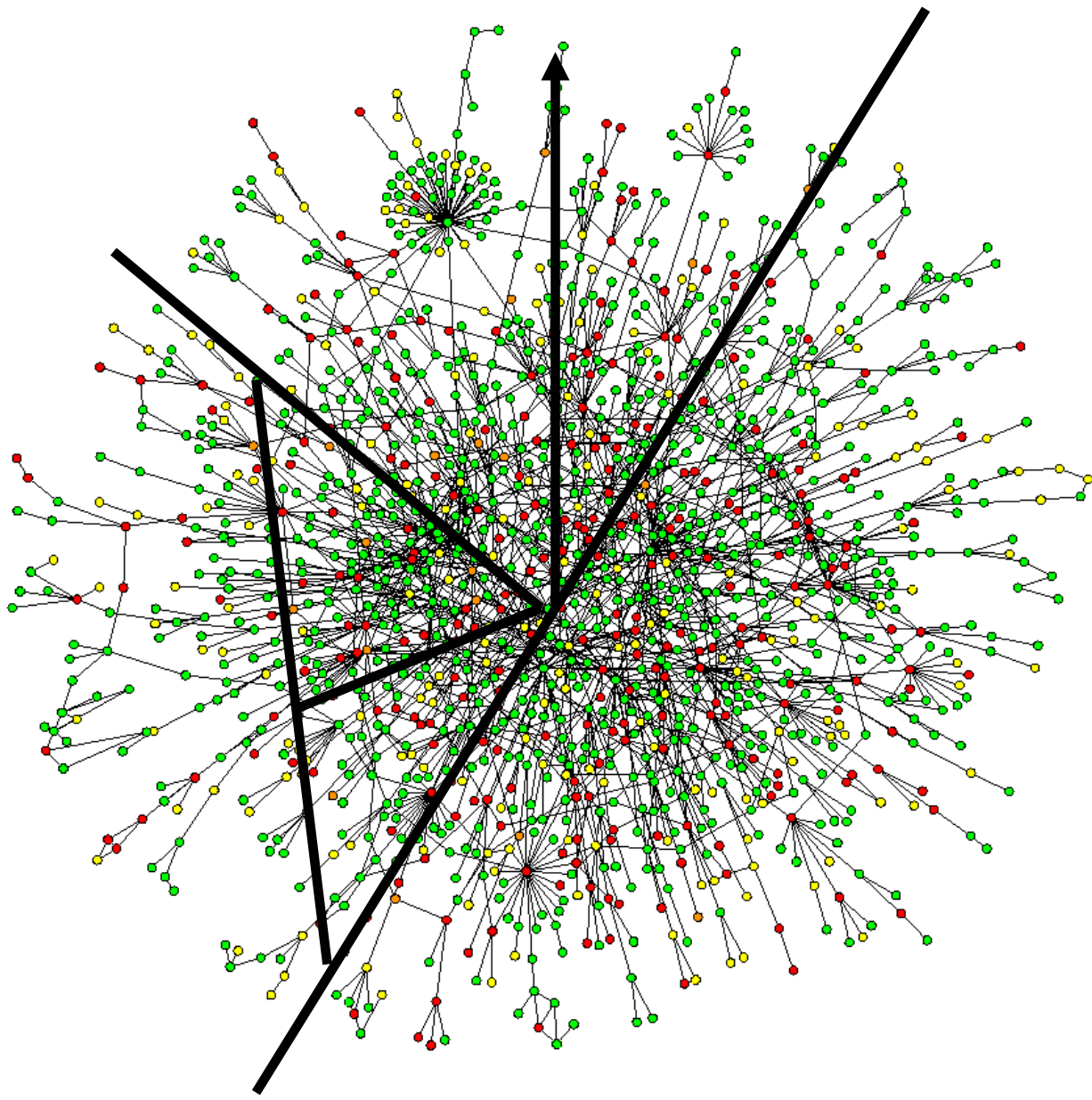
Write the solution like this

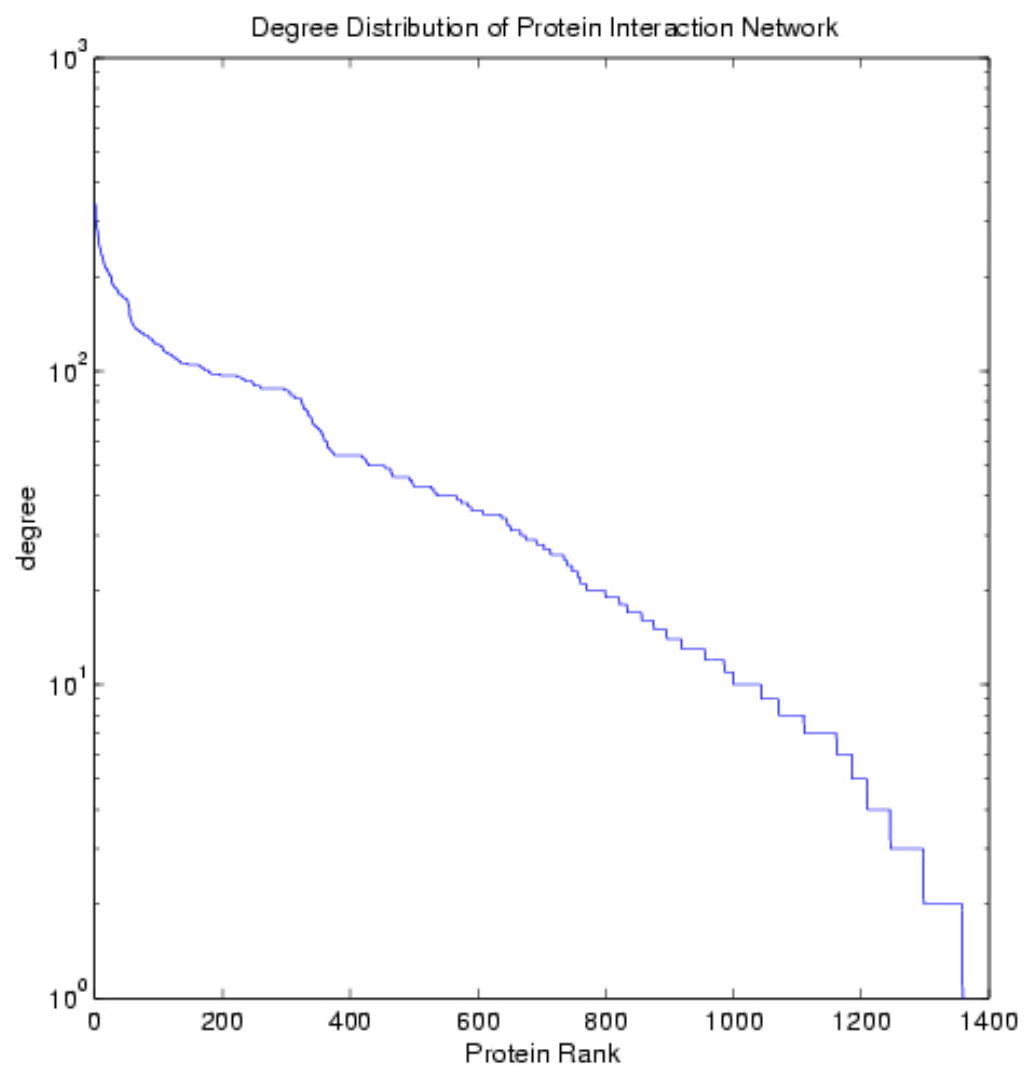
It follows (ie. Proof omitted) :

$$\min_{\mathbf{q}} J(G_1, G_2) \Rightarrow \min_{\mathbf{q}} \frac{\mathbf{q}^T (\mathbf{D} - \mathbf{W}) \mathbf{q}}{\mathbf{q}^T \mathbf{D} \mathbf{q}}$$

Solution is eigenvector  
corresponding to second  
smallest eigenvalue

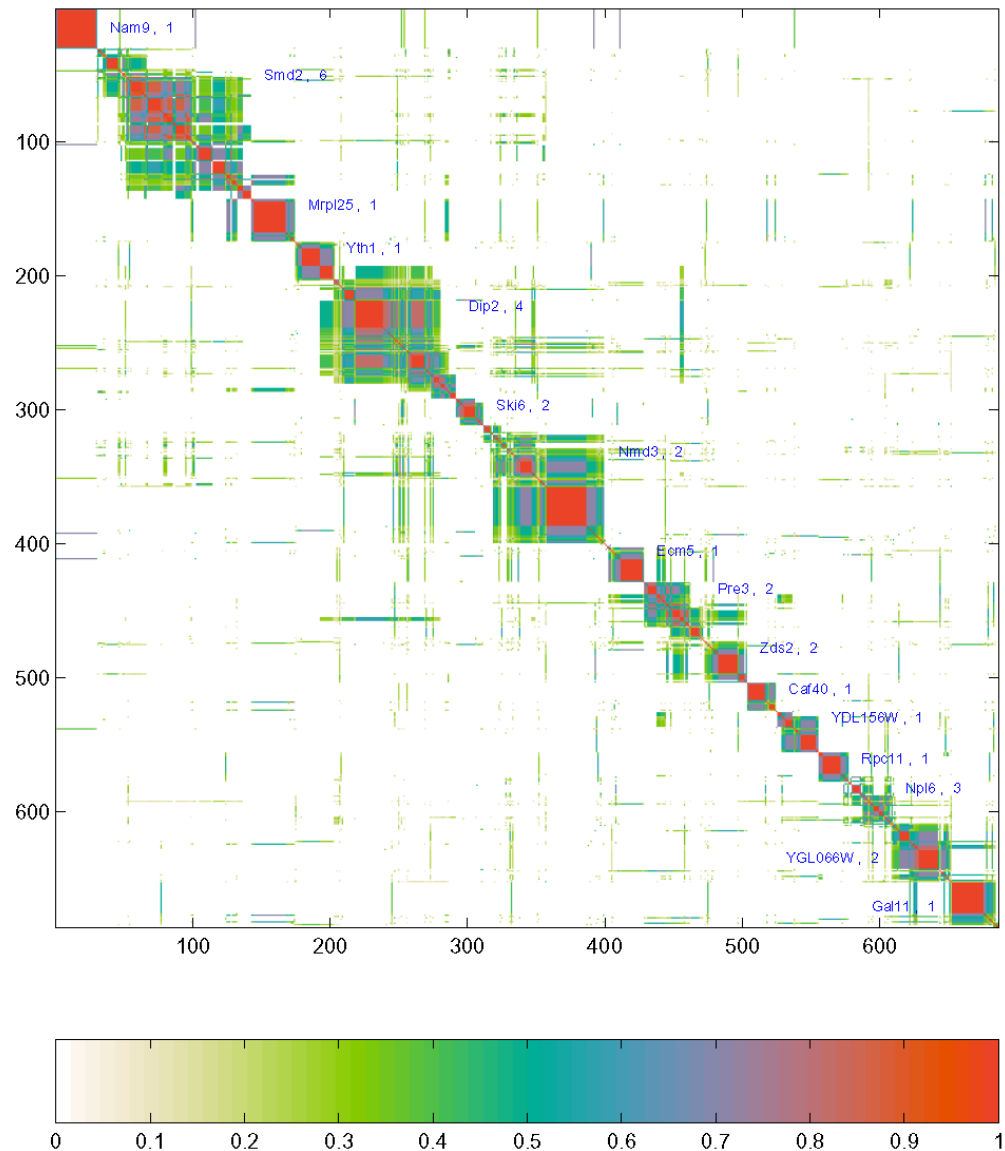
$$(\mathbf{D} - \mathbf{W}) \mathbf{q} = \lambda \mathbf{D} \mathbf{q}$$



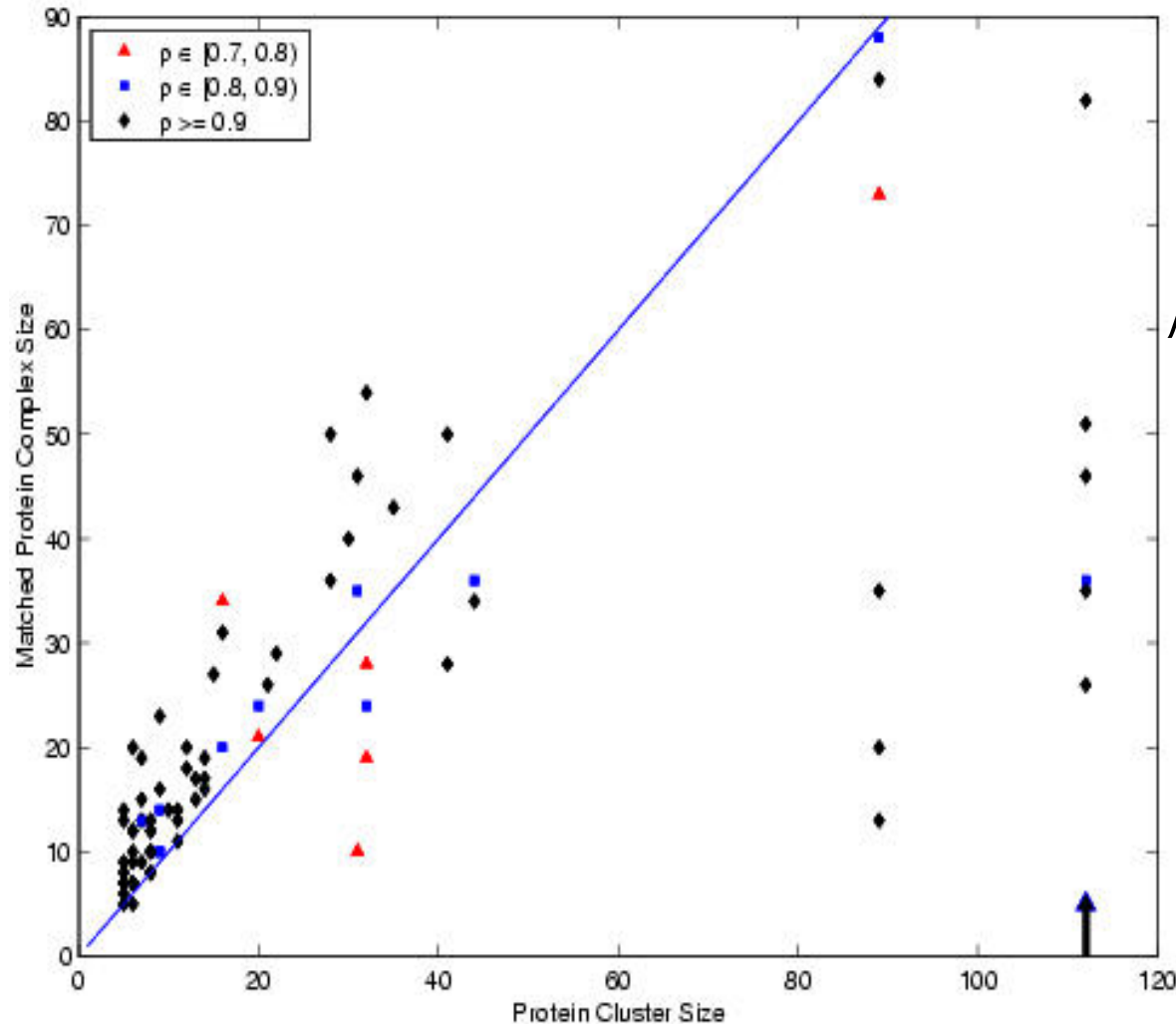


# Predicted Modules(clusters) of P-P network

Connections in the network are weighted by the number of complexes in which two proteins are coincident.



# PP-modules overlap experimental complexes



Overlap between protein clusters and protein complexes defined as

$$\rho = n(P_k, c_j) / \min(|P_k|, |c_j|)$$

- Discovered protein clusters **highly overlap** with experimental complexes
- **Uncharacterized proteins** in discovered clusters might **infer novel functions**.

# Modules in P-P network reflect characteristic physical and chemical properties

Lys	100	Asn	56	Val	30	Ile	24
Asp	89	Gln	50	Tyr	29	Ser	23
Arg	73	Cys	39	Met	29	Leu	22
Pro	70	His	33	Trp	28	Gly	21
Glu	66	Ala	31	Thr	28	Phe	21
pI	169	Basic	149	Acidic	97	MW	60
Aromatic	30	Helix	37	Beta-Sheet	33	Coil	27

$$F = \frac{1}{K-1} \sum_{k=1}^K n_k (\bar{f}_k - \bar{f}) / \frac{1}{n-K} \sum_{k=1}^K (n_k - 1) \sigma_k^2$$

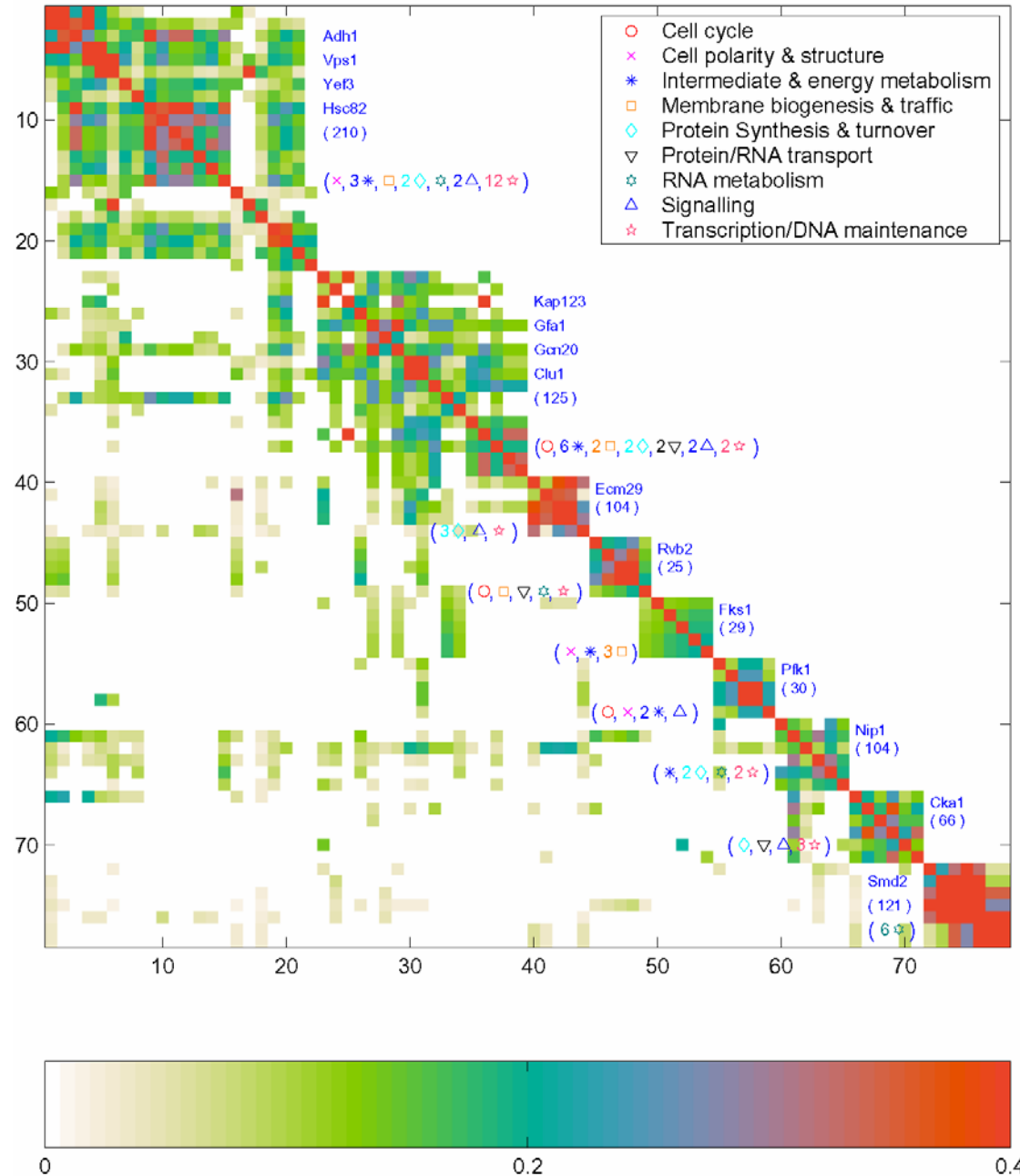
Polar residues (Lys, Arg, Gln, Asn, Asp), hydrogen bonding (Arg), hydrophobic interactions (Pro).

Covalent Modification (methylation and acetylation) of Arg and Lys

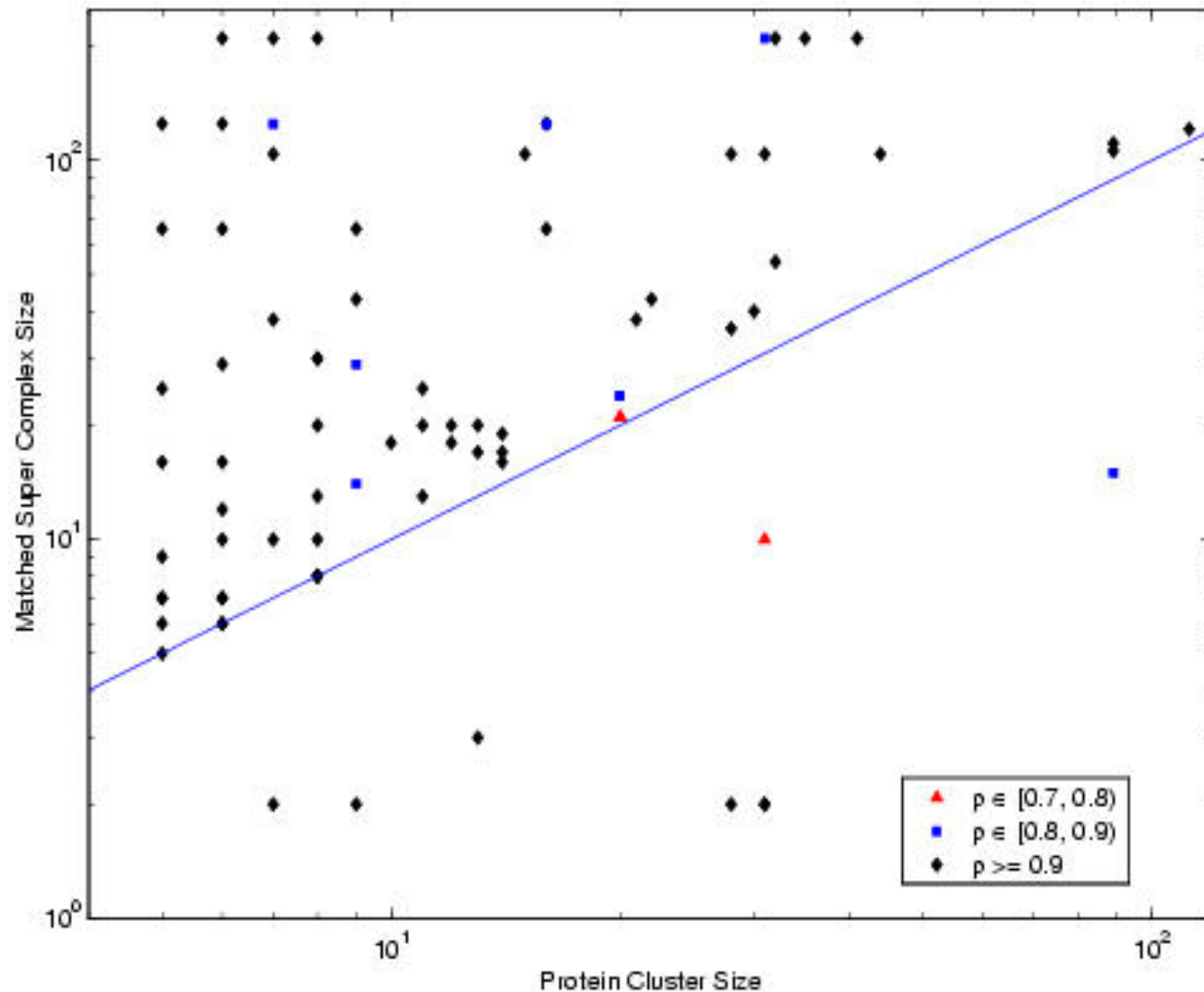
Disulfide bonds and cys.

Secondary structure features uniformly distributed at protein interaction interfaces.

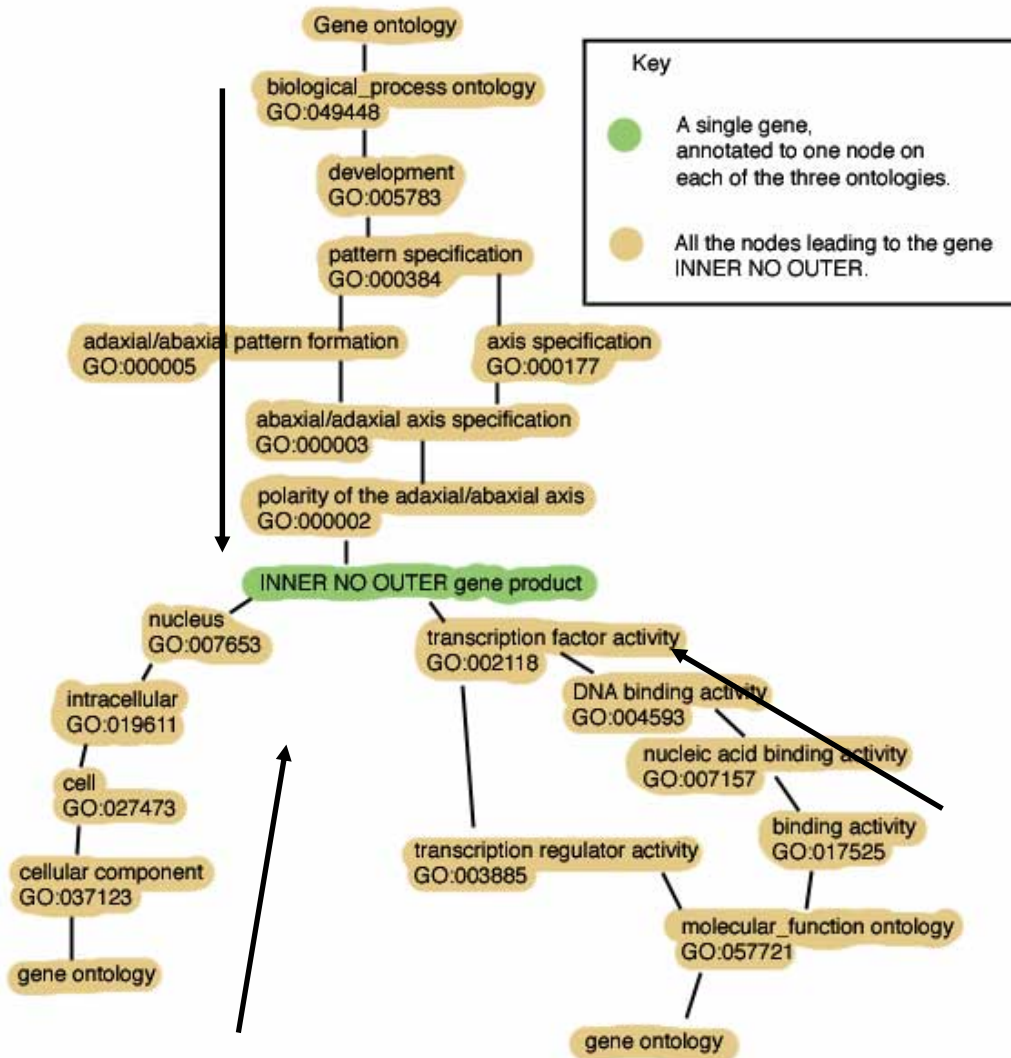
Connections in the network are weighted by the number of proteins that two complexes share.



# Supercomplexes overlap modules in PP-network



# Gene Ontology (GO)



Three separate ontologies:  
**Biological Process**,  
**Molecular Function**, **Cellular Component**.

Organized as a **DAG**  
describing gene products  
(proteins and functional  
RNA).

Makes the represented  
biological relationships  
computable.

Collaborative effort between  
major genome databases.

<http://www.geneontology.org>



# Gene Ontology

- Molecular function **catalytic or binding activities at (e.g. nucleic acid binding or exonuclease)**
- Biological process **is accomplished by ordered assemblies, pathways, with concerted function (e.g. 'signal transduction' or 'nuclear export').**
- Cellular component **compartmental, obligatory, or logical grouping (e.g. nucleus or spliceosome).**



# Validating the Modules in the Networks

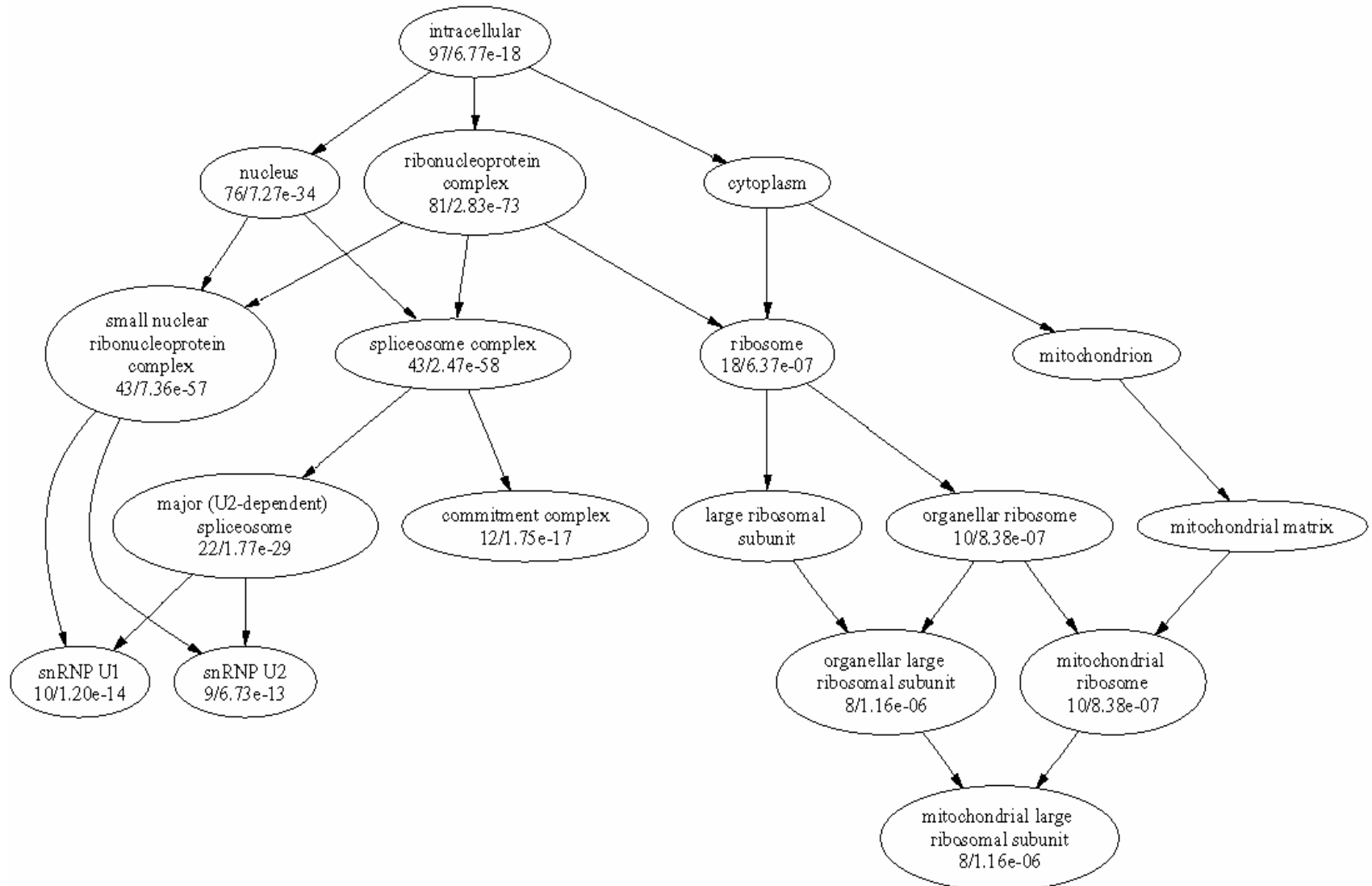
DAG structure of GO formalizes knowledge in biology by making it computable.

For constituent proteins in each cluster annotate  
To most specific term. Ascend the graph and  
annotate with parent terms.

Annotations observed by chance?

$$P = \sum_{n \leq j \leq N} \binom{N}{j} p^j (1-p)^{N-j}$$

# Computationally Discovered Modules are Biologically Consistent





# Verified Complexes in Supercomplex 47

MIPS Annotation Category	# ORFs in $C_{47}$	# ORFs matched
RNA Pol II holoenzyme	35	23
Kornberg's mediator	21	21
Other transcription	73	17
HAT A	15	14
TFIID	13	13
SAGA	14	13
Ada-Spt	14	13
TAFIIIs	12	12
DNA repair	33	9
RSC	10	6
ADA	6	6
Replication fork	30	6
DNA mismatch repair	5	5
Cytoplasmic translation initiation	27	4
SAGA-like	5	4
Nucleotide excision repairosome	16	3
RNA Polymerase III	13	3
Replication factor A	3	3
Actin-associated motorproteins	7	3
MSH2/MSH3	3	3
Srb10p	4	3
NEF4	2	2
eIF4A	2	2
NuA4	2	2
Nuclear pore	24	2
Sir	2	2

- Transcription
- Gene silencing
- Replication
- RNA processing
- RNA modification
- RNA stability
- mRNA translation
- Protein stability
- Protein translocation
- Metabolite sensing and regulation

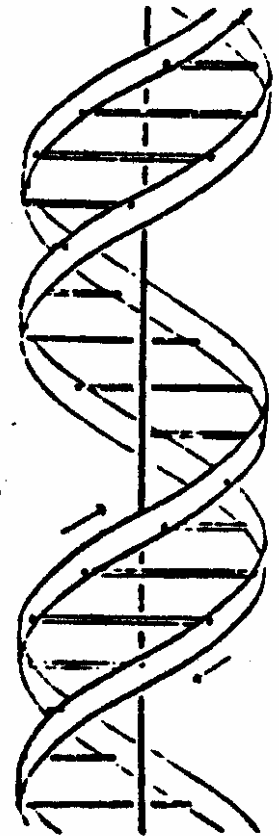
***The Number of Known Functional RNAs  
in E. coli has Grown from 10 to 70  
since 2000.***



# The genome, famously, is digital

1892: Miescher postulates that genetic information may be encoded in a linear form using a few different chemical units:

*“...just as all the words and concepts in all languages can find expression in twenty-four to thirty letters of the alphabet.”*



This figure is purely diagrammatic. The two ribbons symbolize the two phosphate—sugar chains, and the horizontal rods the pairs of bases holding the chains together. The vertical line marks the fibre axis

# Symbolic texts can be cracked

*Michael Ventris and John Chadwick, 1953*



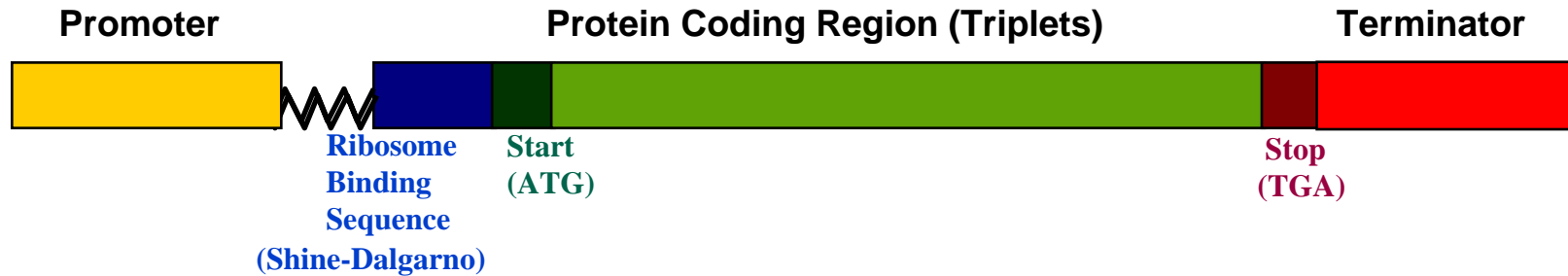
“Cryptography has contributed a new weapon to the student of unknown scripts.... the basic principle is the analysis and indexing of coded texts, so that underlying patterns and regularities can be discovered. *If a number of instances can be collected, it may appear that a certain group of signs in the coded text has a particular function....*”

- John Chadwick,  
*The Decipherment of Linear B*,  
Cambridge Univ. Press, 1958

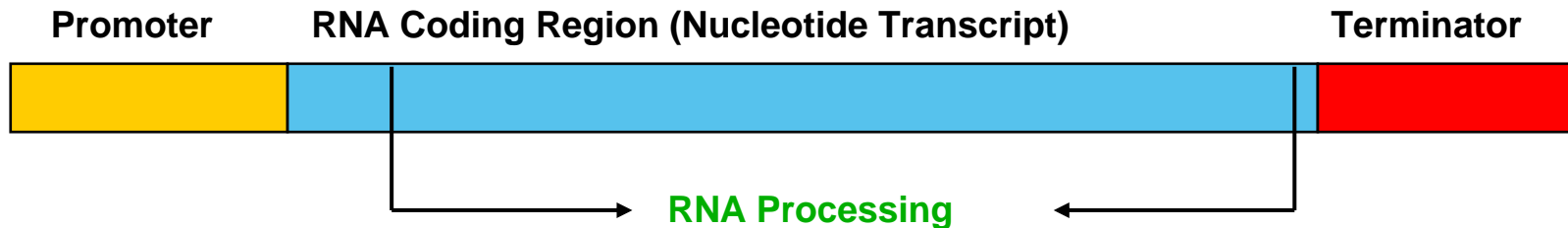


# Microbial Protein and RNA Genes

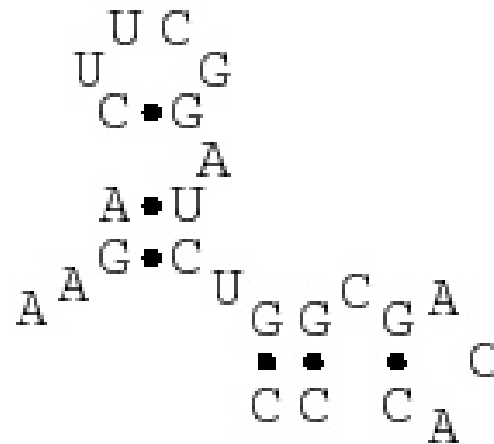
## Protein Gene



## RNA Gene



- No ribosome binding sites
- No start or stop codons
- No triplet code

PHYSICAL BIOSCIENCES DIVISION

# Context-free grammars

*Noam Chomsky, 1956*


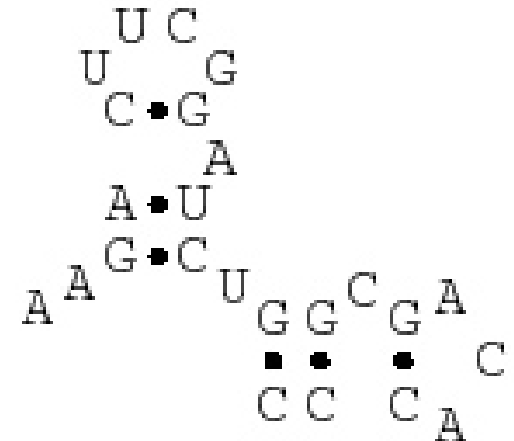
Basic CFG

“production rules”

$S \rightarrow aS$   
 $S \rightarrow Sa$   
 $S \rightarrow aSu$   
 $S \rightarrow SS$

a CFG “derivation”

$S \rightarrow aS$   
 $\rightarrow aaS$   
 $\rightarrow aaSS$   
 $\rightarrow aagScuS$   
 $\rightarrow aagaSucugSc$   
 $\rightarrow aagaSaucggScc$   
 $\rightarrow aagacSgaucuggcScc$   
 $\rightarrow aagacuSgaucuggcgSccc$   
 $\rightarrow aagacuUSgaucuggcgacSccc$   
 $\rightarrow aagacuucSgaucuggcgacSccc$   
 $\rightarrow aagacuucgSgaucuggcgacacSccc$   
 $\rightarrow aagacuucggaucuggcgacacccc$

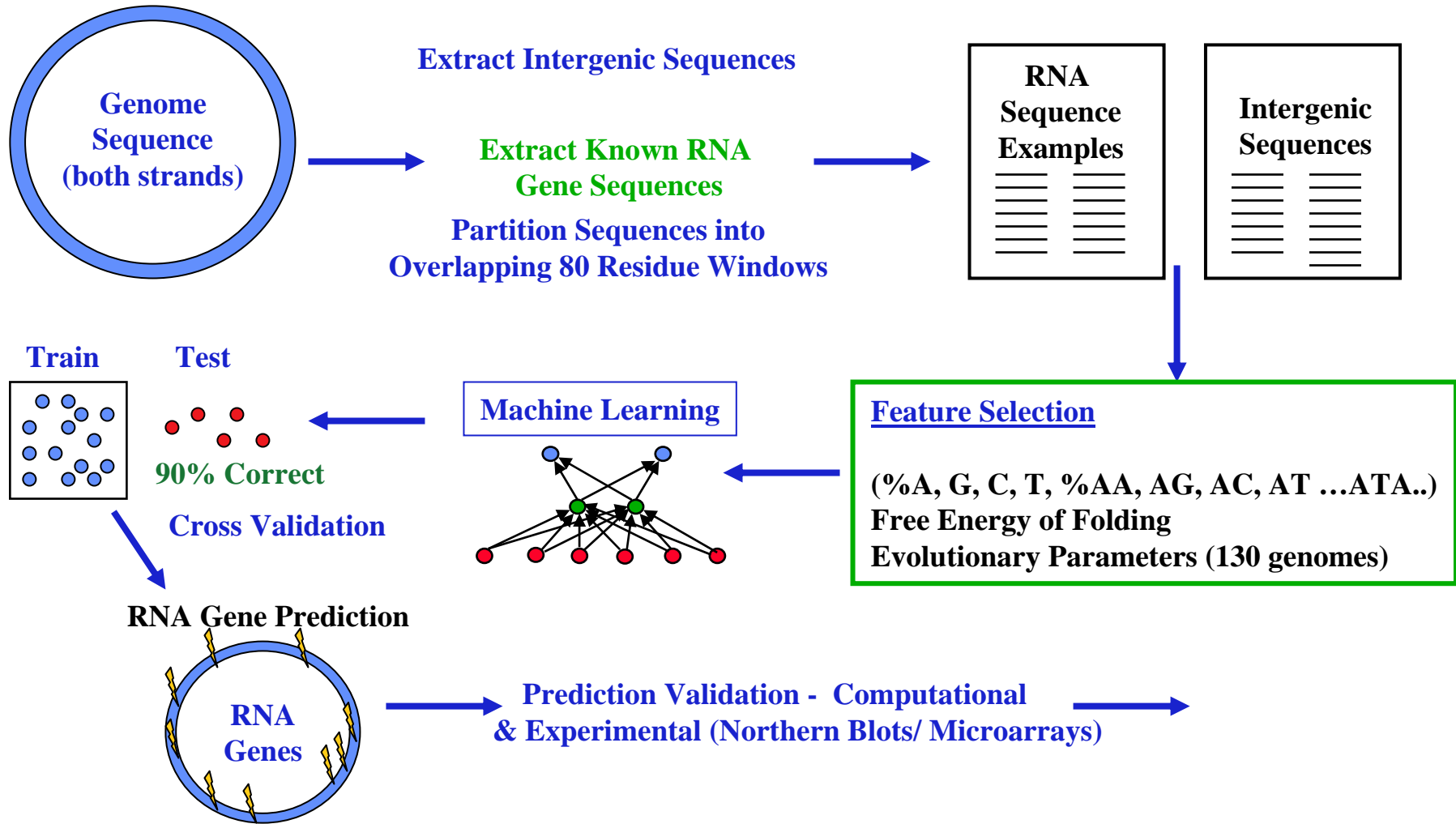


# Machine Learning

## Definition:

**A computer program is said to *learn* from experience  $E$  with respect to some class of tasks  $T$  and performance measure  $P$ , if its performance at tasks in  $T$ , as measured by  $P$ , improves with experience.**

# Flow Chart for RNA Gene Prediction



# Learning from Partially Labeled Data

Determine initial negative set (N) such that :

- (1) Maximally distant from positive set (P)
- (2) Maximally dissimilar from each other

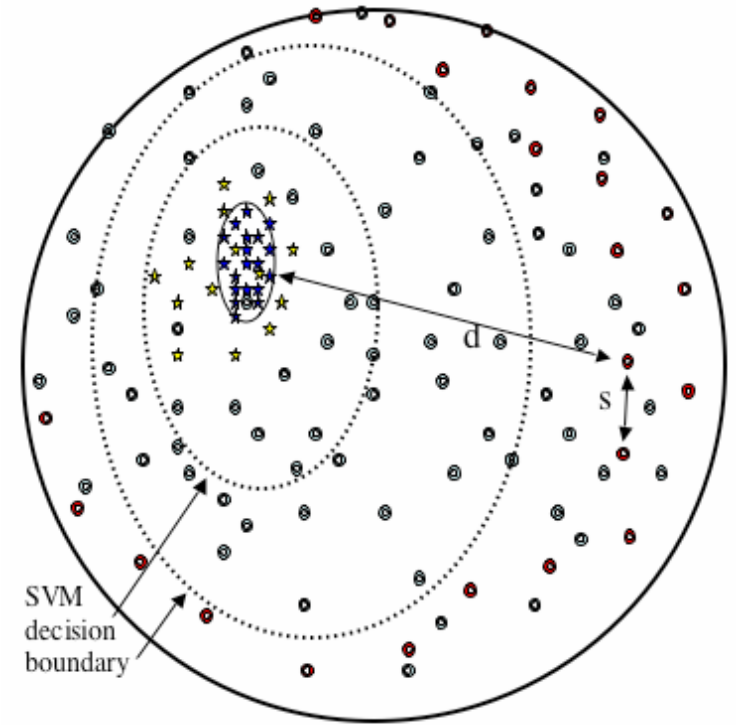
$$(1) \max_{N \subset U} d(N, P), \quad d(N, P) = \sum_{i \in N} d(x_i, P)$$

$$(2) \max_{N \subset U} d(N, N), \quad d(N, N) = \sum_{i, j \in N} d(x_i, x_j)$$

where  $d(x_i, P) = \min_{j \in P} \|x_i - x_j\|$

Solution is messy and expensive for lots of data.

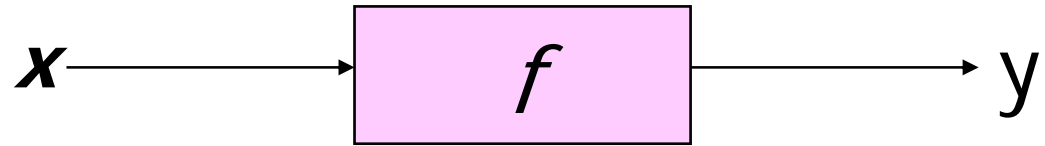
$$\max_{N \subset U} [d(N, P) d(N, N)] \longrightarrow$$



Close enough and easy to compute

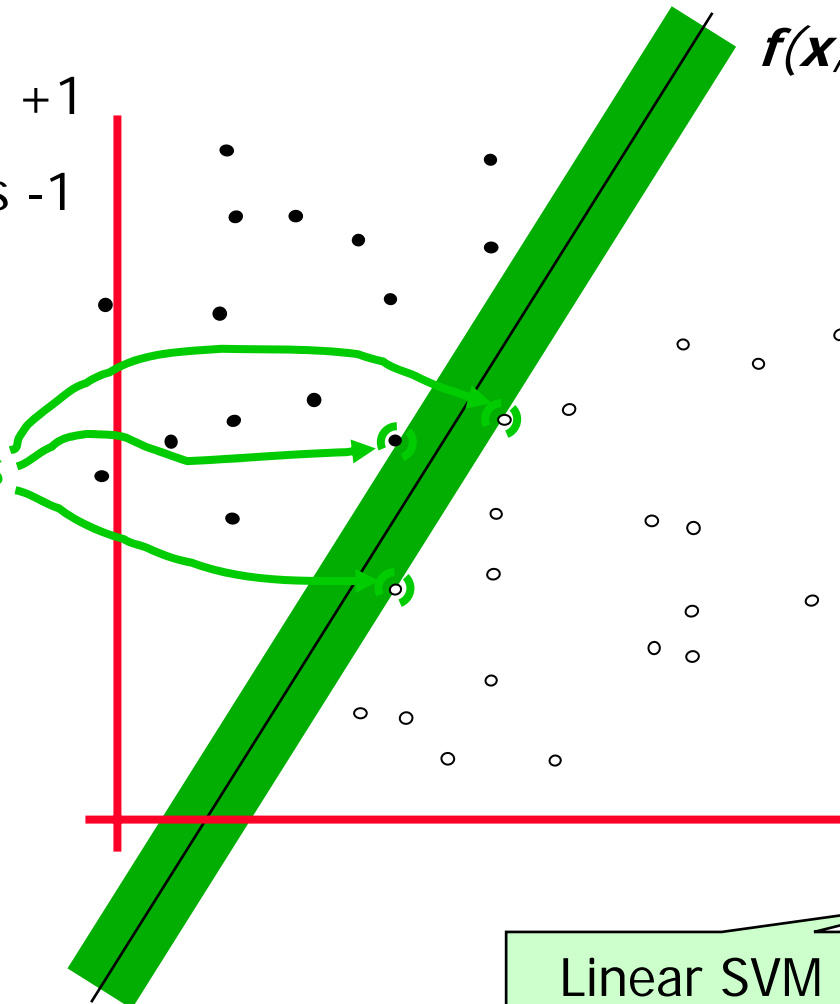
$$\max_{i \in (U-S)} \left[ d(x_i, P) \sum_{j \in S} d(x_i, x_j) \right]$$

# Maximum Margin Hyperplane



- denotes +1
- denotes -1

**Support Vectors**  
are those data points that the margin pushes up against



$$f(x, w, b) = \text{sign}(w \cdot x + b)$$

The **maximum margin linear classifier** is the linear classifier with the, um, maximum margin.

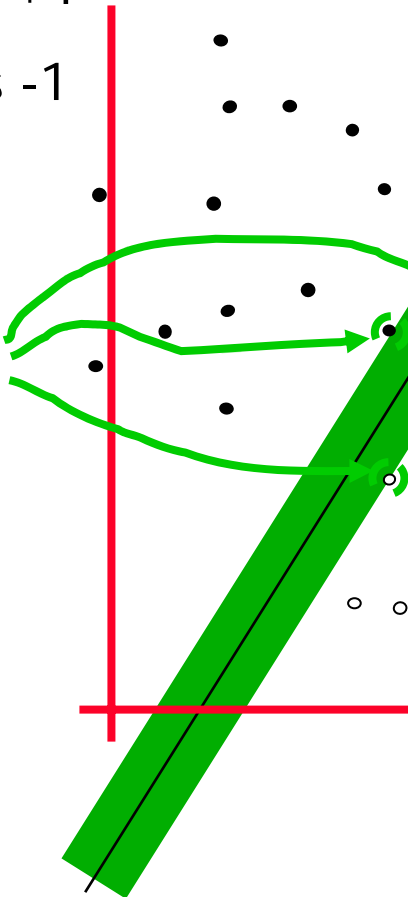
This is the simplest kind of SVM (Called an LSVM)

Linear SVM

# Why Maximum Margin?

- denotes +1
- denotes -1

**Support Vectors**  
are those data points that the margin pushes up against



1. Intuitively this feels safest
2. If we've made a small error in the location of the boundary (it's been jolted in its perpendicular direction) this gives us least chance of causing a misclassification
3. Robust to outliers since the model is immune to change/removal of any non-support-vector data points
4. There's some theory (using VC dimension) that is related to (but not the same as) the proposition that this is a good thing
5. Empirically it works very well



# SVM Kernel Functions

- $K(a,b)=(a \cdot b +1)^d$  is an example of an SVM kernel function
- Beyond polynomials there are other very high dimensional basis functions that can be made practical by finding the right kernel function

—Radial-Basis-style Kernel Function:

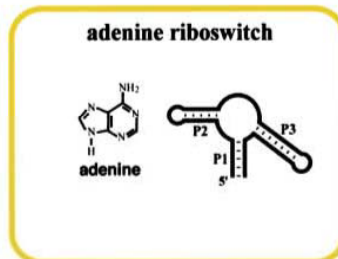
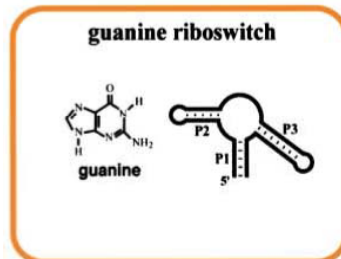
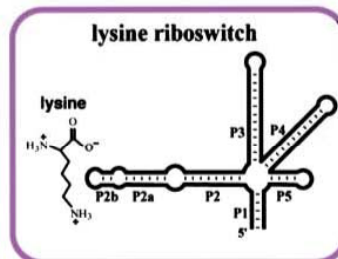
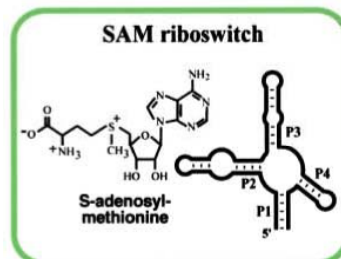
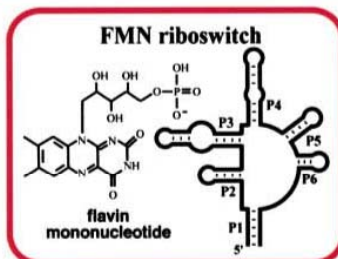
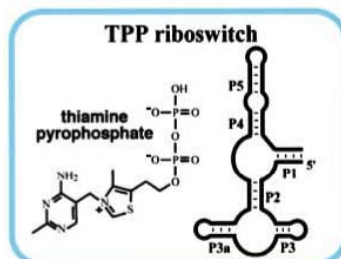
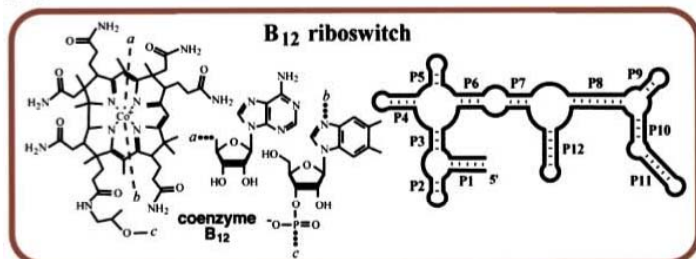
$$K(\mathbf{a}, \mathbf{b}) = \exp\left(-\frac{\|\mathbf{a} - \mathbf{b}\|^2}{2\sigma^2}\right)$$

—Sigmoid Style Kernel Function:

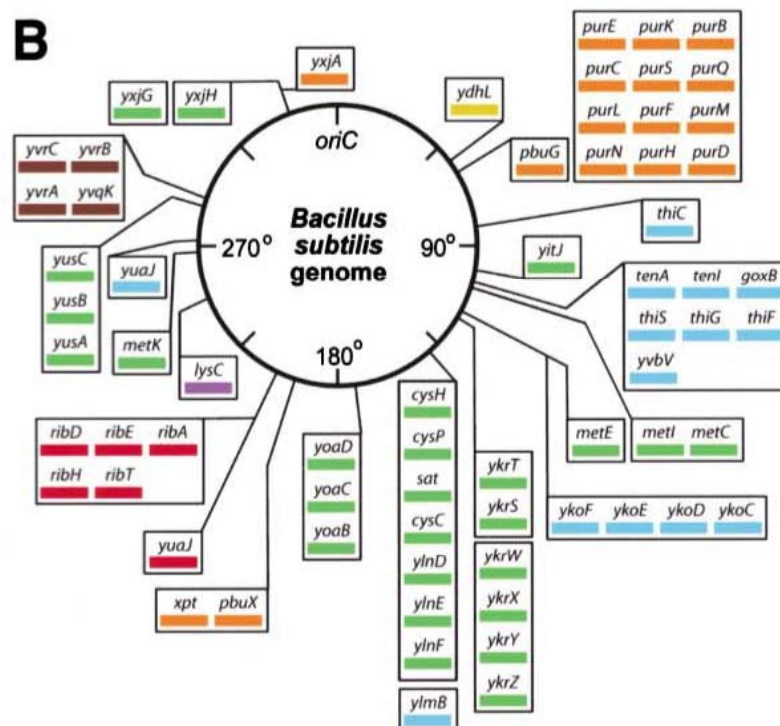
$$K(\mathbf{a}, \mathbf{b}) = \tanh(\kappa \mathbf{a} \cdot \mathbf{b} - \delta)$$

$\sigma$ ,  $\kappa$  and  $\delta$  are magic parameters that must be chosen by a model selection method such as CV or VCSRM

**A**



**B**



## RIBOSWITCHES IN FUNDAMENTAL GENE CONTROL

A. THE SEVEN KNOWN RIBOSWITCHES AND THE METABOLITES THEY SENSE; NOTE THAT THE METABOLITES ALMOST ALL CONTAIN PYRIMIDINE OR PURINE MOIETIES.

B. GENETIC MAP OF *Bacillus subtilis* RIBOSWITCH REGULONS AND THEIR POSITIONS ON THE BACTERIAL CHROMOSOME; GENES ARE CONTROLLED BY RIBOSWITCHES OF MATCHING COLOR.

# Structure of the large ribosomal subunit

*Haloarcula marismortui*

*Ban et. al., Science 289:905, 2000*

